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Attorney's Docket No. 080743-235001

PATENT
TECH CENTER 1600/2900

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of: VAN DEN BERG Group No.: 1645

Serial No.: 10/ 007,275

Filed: 10/26/2001

Examiner:

For: Method for Inhibiting Cell Functioning for Use in Anti-inflammatory
and Anti-tumor Therapies

Commissioner of Patents and Trademarks
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Country : European Patent Application (EP)

Application Number: 99201350.8

Filing Date : April 28, 1999

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Method for inhibiting cell functioning for use in anti-inflammatory and anti-tumour therapies

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METHOD FOR INHIBITING CELL FUNCTIONING FOR USE IN ANTI-INFLAMMATORY AND ANTI-TUMOUR THERAPIES

5 The invention relates to a method for inhibiting cell functioning for use in anti-inflammatory and anti-tumour therapies in the body of a warm-blooded living being. The invention further relates to a drug to be used in the above method, and to the active substance of said drug.

10 Inflammations in the body of a warm-blooded living being, in particular a human being, cause many diseases and disorders, and may even turn out to be life-threatening. Therefore, for many decades already it is a major challenge to the clinician to find an effective therapy in treating
15 inflammatory diseases. Various inflammatory diseases, such as rheumatoid arthritis, multiple sclerosis, glomerulonephritis, diabetes and asthma, are the result of unwanted immune responses. As described, for instance, in a recent survey entitled "Manipulation of the Immune
20 Response" ("Immunobiology", 3rd Edition; C.A. Janeway, P. Travers; publ. Current Biology/ Garland/ Churchill Livingstone 1997; Chapter 13), current treatments for immunological disorders are nearly all empirical in origin using immunosuppressive drugs identified by screening large
25 numbers of natural and synthetic compounds.

According to this survey, these drugs may be divided into three categories, viz. (i) drugs of the corticosteroid family, (ii) cytostatic drugs, and (iii) fungal and bacterial derivatives. In this survey it is noted, that
30 these drugs are all very broad in their actions and inhibit protective functions of the immune system as well as harmful ones. In fact, the ideal immunosuppressive agent would be a drug that targets the specific part of the immune response responsible for causing the relevant tissue injury.
35 Consequently, according to this survey, antibodies themselves, by virtue of their exquisite specificity, may offer the best possibility for the therapeutic inhibition of specific immune responses. Such immunosuppressive

monoclonal antibodies can act by inhibiting target cell functioning. Their promising potential in immunosuppression has already been established. However, as yet these antibodies are not widely and generally used as anti-inflammatory drugs, largely due to the fact that the appropriate targets have not been identified.

It is the objective of the present invention to provide a method for inhibiting or controlling target cell functioning, for use in anti-inflammatory and anti-tumour therapies in the body of a warm-blooded living being by administration of a drug, having superior therapeutic properties compared to existing anti-inflammatory and anti-tumour drugs. Various requirements should be imposed on a drug to be used in such therapies, for example, non-toxic, no adverse influence on the host resistance, and highly selective to avoid burdening of non-target tissues and organs with drug material.

According to the present invention the above-defined objective can be achieved by a method which comprises administering to said being a drug comprising, in a quantity effective for said therapies, a substance that specifically recognizes the extracellular domain of SIRP (= signal regulatory protein) (anti-SIRP substance) and that inhibits the functioning of pathologic myeloid cells. By using a drug according to the method of the present invention, both a highly selective and an effective therapy in treating inflammatory diseases, in particular autoimmune diseases and allergies, and tumours can be achieved.

Well-known examples of myeloid cells are macrophages, which are continuously replenished from a population of dividing and maturing myeloid precursor cells in the bone marrow. This ensures the continuous availability of macrophages in all tissues of the body and allows a fast and efficient response in case of infections. In a number of

circumstances, however, macrophages do not play a beneficial role. In a variety of autoimmune diseases, like rheumatoid arthritis, multiple sclerosis, glomerulonephritis etc., and allergies, like asthma, activated macrophages play an important role in the induction and/or maintenance of inflammations that, as a consequence, forms the basis for the (general chronic) clinical symptoms.

In addition, under certain circumstances the myeloid precursor cells may cause pathologies; the unlimited growth of these myeloid precursor cells is the cause of certain malignant tumours, in particular myeloid leukemia.

More in particular, said anti-SIRP substance to be used in the method of the present invention is characterized in that it inhibits the functioning of macrophages by suppressing their activation by a factor of at least 10 as measured by each of the following so-called macrophage activity tests: (i) the production of nitric oxide (NO), (ii) the production of reactive oxygen species, in particular superoxides (e.g. H_2O_2), and (iii) the production of tumour necrosis factor - alpha (TNF).

The above tests for measuring the activity of macrophages are described in detail in Example II hereinafter. It has been found, that the substances to be used according to the method of the invention show the above striking effect in all three above tests. Therefore these tests are a convenient tool of distinguishing substances within the scope of the invention from other compounds.

Signal-regulatory proteins (SIRP), as recently disclosed by S. Adams et al. in J. Immunol. 161: 1853-1859 (1998), are transmembrane glycoproteins, inhibiting signaling through receptor tyrosine kinases but having a physiological function which is unknown up to the present. SIRP is selectively expressed on the surface of myeloid cells, such as macrophages, monocytes, granulocytes and dendritic cells, and neurons. The active substances to be used in the method

5 following requirements:

- 10 to the three tests as described above.

15 a malignant tumour. It has been found that the above anti-SIRP substances to be used according to the method of the invention can also inhibit the functioning of pathologic myeloid cells by strongly suppressing the division of macrophage tumour cell lines. More specifically, the
20 suppression of this cell division is also found to amount to a factor of at least 10, as demonstrated in the so-called macrophage division test (Examples II).

25 tumour therapy, in particular for treating myeloid leukemia,
because the selective binding of these substances to the
extracellular domain of SIRP can effectively and selectively
control the division of myeloid cells.

30 The above-mentioned functioning of myeloid cells, in particular macrophages, encompasses not only their activation and division, but also the phenomenon of phagocytosis, that is the uptake of other organisms or other particles.

35 In case of gene-targeted therapies, e.g. gene-targeted anti-tumour therapy, where genes packed in vector particles (vehicles) are targeted to different cells or tissues,

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modified products of such fragments wherein the intended anti-SIRP activity has been maintained.

The invention will now be described in greater detail with
5 reference to the following specific Examples.

Example I

Preparation of Fab-fragments of ED9 and ED17:

10 The starting monoclonal antibodies ED9 and ED17 are disclosed by Damoiseaux et al. in J. Leukocyte Biol. 46:556-564 (1989) and 49: 434-441 (1991). The Fab-fragments of these antibodies are obtained by papain-protolytic digestion. For this purpose a papain-solution, containing
15 0.1 mg of papain per ml PBS buffer solution (0.02M EDTA and 0.02M cystein in PBS), is added to the same volume of a solution of the antibody (1 mg/ml) in PBS. The mixture is incubated at 37°C, and after a certain time, determined by making a time-series, the reaction is stopped by adding a
20 0.03M iodoacetamide solution (addition of 20 μ l 0.3M iodoacetamide to 110 μ l incubated mixture). The mixture is now dialysed against 2 l PBS at pH 8.0, O/N at 4°C. The solution is chromatographed over a protein A sepharose column, concentrated to 5 ml at reduced pressure, and
25 chromatographed over a superose 12 column. The fractions of 50 kD are received and purity-controlled on non-reduced SDS-PAGE^R. The solution of the Fab-fragments ED9 and ED17, so obtained, are used as such in the cell culture experiments described in Example II.

30

Example II

Cell culture experiments

The macrophage activity test

35 Rat peritoneal macrophages, obtained by peritoneal lavage, of the rat macrophage cell line NR8383 (Adams et al. 1998) are cultured at a density of 0.25×10^6 cells/ml in RPMI-

1640 medium containing 2% fetal calf serum and 2 mM glutamine, 100 U/ml penicillin and 100 µg/ml streptomycin. Macrophage activating stimuli (100 ng/ml lipopolysaccharide (LPS), or 20 U/ml gamma-interferon (IFN)-γ) are added in the presence (or absence) of anti-SIRP Fab-fragments (ED9 or ED17; 40 µg/ml) or control Fab-fragments (OX41, Adams et al. 1998; 40 µg/ml). After 18-20 hours the cell culture supernatants (separated from the cells by centrifugation for 7 min. at 500g) are harvested. NO production in supernatants is measured using Griess reagent (Ding et al. (1988), J. Immunol. 141:2407) using NaNO₂ to produce a calibration curve. TNFα, IL1β and IL6 are measured by enzyme-linked immunosorbent assay as described (Vincent et al. (1996), Glia 17:94; Lenczowski et al. (1997), Am.J.Physiol. 273:R1870). The results are presented in the diagram of Figure 1.

The macrophage phagocytosis test

0.5 x 10⁶ rat peritoneal macrophages are plated in each well of a 24-well cell culture plate in RPMI-1640 medium containing 10% fetal calf serum and 2 mM glutamine, 100 U/ml penicillin and 100 µg/ml streptomycin, and are then allowed to adhere for 1-1.5 hours at 37°C in a 5% CO₂ atmosphere. After this the cells are washed 2x and incubated with 0.5 ml HEPES (25 mM)-buffered RPMI containing 2 µg oxidated LDL (low-density lipoproteins), 2 µg acetylated LDL (both: FITC-labelled; Molecular Probes), 1 µl latex beads (FITC-labelled; Molecular Probes), 2 µg serum treated zymosan (FITC-labelled), or rat myelin (DiI-labelled) plus 5% fresh rat serum. These incubations are performed in the presence (or absence) of anti-SIRP Fab-fragments (ED9 or ED17; 40 µg/ml) or control Fab-fragments (OX41; 40 µg/ml). After 1.5 hours the cells are washed to remove non-bound particles, cells are detached by incubation in 5 mM EDTA in PBS and mean fluorescence intensity for each cell is measured on a FACScan[®]. Values are plotted as the percentage of control phagocytosis: Figure 2.

FITC and DiI are fluorescent dyes, well-known in the art.

The macrophage division test

The rat macrophage cell line NR8383 (Adams et al. 1998) are
 5 cultured at a density of 0.25×10^6 cells/ml in a 96-well
 cell culture plate in RPMI-1640 medium containing 2% fetal
 calf serum and 2 mM glutamine, 100 U/ml penicillin and 100
 µg/ml streptomycin. This is performed in the presence (or
 absence) of anti-SIRP Fab-fragments (ED9 or ED17; 40 µg/ml)
 10 or control Fab-fragments (OX41; 40 µg/ml). After 24 h ^3H -
 thymidine (1 µCi/well) is added and the cells are incubated
 for another 6 hours. The cells are harvested using a cell
 harvester and cell incorporated radioactivity is determined
 in a Micro-β-plate reader. The mean results are shown in
 15 Table 1 below:

Treatment	Mean (in c.p.m.)	SD (standard dev.)
control	132783	2730
ED17 Fab	6845	197
20 OX41 Fab	154889	8528

Results

In all above experiments the results of ED9 Fab and of ED17
 Fab are comparable with each other; therefore the results
 25 presented are confined to one active substance.

To evaluate the effects of ED9 or ED17 Fab-fragments, cell
 culture experiments using animal cells, predictive for human
 myeloid and/or inflammatory cells, are performed. In the
macrophage activity test (Figure 1) the effect on the
 30 production of the inflammatory mediators reactive oxygen
 species (H_2O_2 as ROS), nitric oxide (NO) and the
 proinflammatory cytokine $\text{TNF}\alpha$ is measured. As can be seen,
 ED9 Fab strongly suppresses the production of ROS(not
 shown), NO and $\text{TNF}\alpha$, whereas irrelevant OX41 Fab-fragments
 35 do not have this effect.

To evaluate the effect of ED9 or ED17 Fab on phagocytosis
 peritoneal macrophages are assayed as described in the

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Discussion and Conclusion

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CLAIMS

1. A method for inhibiting cell functioning for use in
5 anti-inflammatory and anti-tumour therapies in the
body of a warm-blooded living being, which comprises
administering to said being a drug comprising, in a
quantity effective for said therapies, a substance
10 that specifically recognizes the extracellular domain
of SIRP (anti-SIRP substance) and that inhibits the
functioning of pathologic myeloid cells.
2. The method as claimed in claim 1, wherein said
15 substance inhibits the functioning of macrophages by
suppressing their activation by a factor of at least
10 as measured by each of the following macrophage
activity tests: (i) the production of nitric oxide
(NO), (ii) the production of reactive oxygen species,
20 and (iii) the production of tumour necrosis factor -
alpha (TNF- α).
3. The method as claimed in claim 1, wherein said
substance inhibits the functioning of pathologic
myeloid cells by suppressing the division of
25 macrophage tumour cell lines by a factor of at least
10 as measured by the macrophage division test.
4. The method as claimed in any of claims 1-3 for
30 treating pathologies selected from inflammations,
caused by autoimmune diseases or by allergies, and
myeloid leukemia.
5. The method as claimed in claim 1, wherein said
35 substance inhibits the functioning of macrophages by
temporally suppressing their phagocytosis as measured
by the macrophage phagocytosis test.

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6. The method as claimed in claim 5 for improving the efficacy of gene-targeted therapies.
7. The method as claimed in any of the preceding claims, characterized in that said anti-SIRP substance is selected from the group consisting of Fab-fragments of monoclonal antibodies and (bio)chemically modified products of such fragments wherein the intended anti-SIRP activity has been maintained.
8. The method as claimed in claim 7, wherein said anti-SIRP substance is a Fab-fragment of monoclonal antibody ED9 or ED17, or said modified product thereof.
9. Use of a substance, that specifically recognizes the extracellular domain of SIRP (anti-SIRP substance) and that inhibits the functioning of pathologic myeloid cells, for the manufacture of a drug for inhibiting cell functioning for use in anti-inflammatory and anti-tumour therapies.
10. The use as claimed in claim 9, wherein the anti-SIRP substance is selected from the group consisting of Fab-fragments of monoclonal antibodies, preferably of ED9 or ED17, and (bio)chemically modified products of such fragments wherein the intended anti-SIRP activity has been maintained.
11. A drug to be used according to any of the preceding claims, comprising, in addition to a pharmaceutically acceptable carrier and, if desired, one or more pharmaceutically acceptable adjuvants, as the active substance an anti-SIRP substance that inhibits the functioning of pathologic myeloid cells.
12. A drug as claimed in claim 11, wherein the anti-SIRP

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substance is selected from the group consisting of Fab-fragments of monoclonal antibodies, preferably of ED9 or ED17, and (bio)chemically modified products of such fragments wherein the intended anti-SIRP activity has been maintained.

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13. An anti-SIRP substance that inhibits the functioning of pathologic myeloid cells, selected from the group consisting of Fab-fragments of monoclonal antibodies, preferably of ED9 or ED17, and (bio)chemically modified products of such fragments wherein the intended anti-SIRP activity has been maintained.

10

The invention relates to a method for inhibiting cell functioning for use in anti-inflammatory and anti-tumour therapies in the body of a warm-blooded living being, which comprises administering to said being a drug comprising, in a quantity effective for said therapies, a substance that specifically recognizes the extracellular domain of SIRP (anti-SIRP substance) and that inhibits the functioning of pathologic myeloid cells.

The invention further relates to a drug to be used in the above method, and to the active substance of said drug.

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Figure 1.

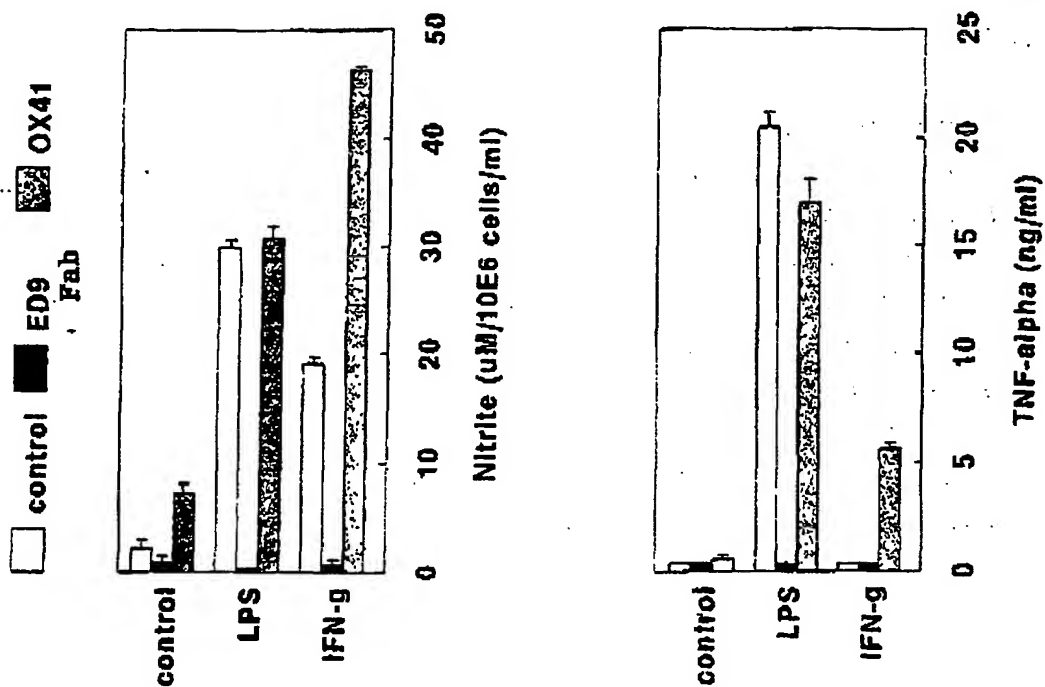
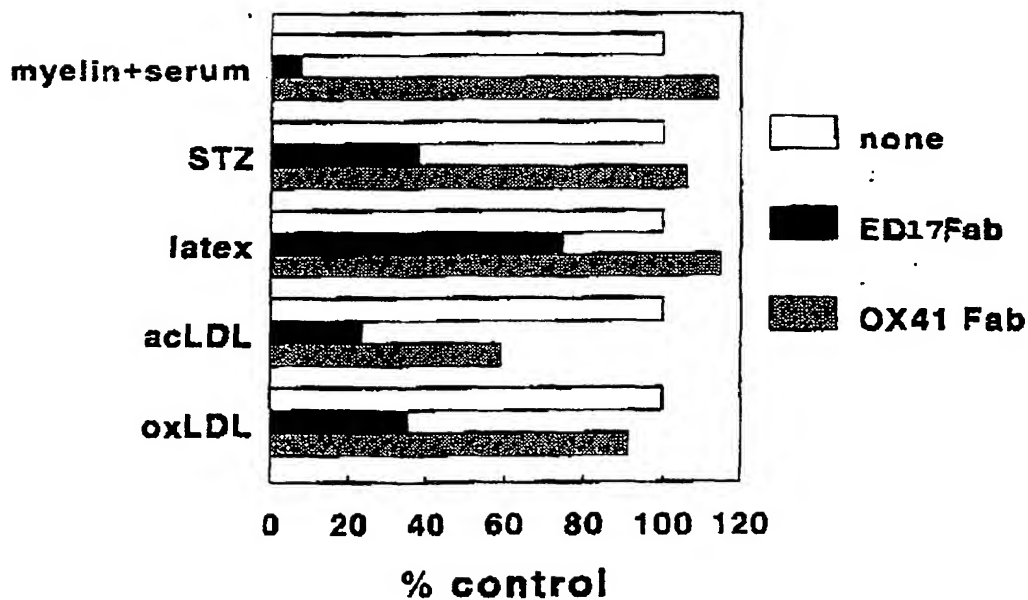


Figure 2.



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